

Themed Section: Pharmacology of the Gasotransmitters

REVIEW

Carbon monoxide and the CNS: challenges and achievements

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Haem oxygenase (HO) and its product carbon monoxide (CO) are associated with cytoprotection and maintenance of homeostasis in several different organs and tissues. This review focuses upon the role of exogenous and endogenous CO (via HO activity and expression) in various CNS pathologies, based upon data from experimental models, as well as from some clinical data on human patients. The pathophysiological conditions reviewed are cerebral ischaemia, chronic neurodegenerative diseases (Alzheimer's and Parkinson's diseases), multiple sclerosis and pain. Among these pathophysiological conditions, a variety of cellular mechanisms and processes are considered, namely cytoprotection, cell death, inflammation, cell metabolism, cellular redox responses and vasomodulation, as well as the different targeted neural cells. Finally, novel potential methods and strategies for delivering exogenous CO as a drug are discussed, particularly approaches based upon CO-releasing molecules, their limitations and challenges. The diagnostic and prognostic value of HO expression in clinical use for brain pathologies is also addressed.

LINKED ARTICLES

This article is part of a themed section on Pharmacology of the Gasotransmitters. To view the other articles in this section visit http://dx.doi.org/10.1111/bph.2015.172.issue-6

Abbreviations

AD, Alzheimer's disease; BBB, blood–brain barrier; BK_{Ca}, large-conductance Ca²⁺-activated K⁺ channels; CMVEC, cerebral microvascular endothelial cell; CORM, carbon monoxide-releasing molecule; DN, dopaminergic neurone; EAE, experimental autoimmune encephalomyelitis; HO, haem oxygenase; IPC, ischaemic pre-conditioning; MCAO, middle cerebral artery occlusion; MPP, 1-methyl-4-phenylpyridinium; MS, multiple sclerosis; PD, Parkinson's disease; ROS, reactive oxygen species; sGC, soluble guanylyl cyclase

Historical aspects

Carbon monoxide (CO) is commonly considered to be toxic because of its high affinity for haem proteins, which can compromise oxygen delivery in tissues, via formation of carboxyhaemoglobin (Bernard, 1857; Haldane, 1895). Claude Bernard was the first to publish an accurate description of the physiology of CO poisoning (Bernard, 1857). About one

century later, CO was also described as cytotoxic by limiting oxidative phosphorylation in cells, via the inhibition of cytochrome *c* oxidase (Wainio and Greenless, 1960; Savolainen *et al.*, 1980; see also Piantadosi, 2002).

Later, CO was recognized as an endogenous molecule in 1949, when Sjostrand (1949) identified this gas as a natural metabolite in the exhaled air of healthy humans. Nevertheless, it was only in 1968 that CO was identified as a product

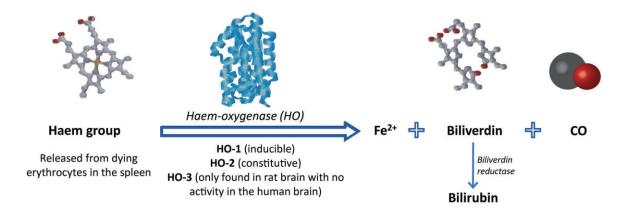


Figure 1

The enzyme haem oxygenase (HO) metabolizes the haem group, giving rise to free iron, biliverdin (rapidly converted to bilirubin) and carbon monoxide.

of haem catabolism by haem oxygenase (HO) (Tenhunen *et al.*, 1968; enzyme nomenclature follows Alexander *et al.*, 2013a), along with free iron and biliverdin (Figure 1).

In 1988, Harbin and colleagues assessed the neurophysiological effects of CO exposure, concluding that acute and low levels of CO exposure were not neurotoxic in normal healthy men (Harbin et al., 1988). In 1993,, CO was accepted as a signalling molecule, being considered a neurotransmitter (Verma et al., 1993). In the beginning of the new millennium, the first therapeutic actions of CO, as a vasomodulator (Sammut et al., 1998), as an anti-inflammatory (Otterbein et al., 2000) and as an anti-apoptotic factor (Brouard et al., 2000) were described. Since then, several distinct applications of CO have been explored, namely in organ transplantation, as a cardioprotective, anti-inflammatory and anti-apoptotic molecule, and for limiting cell proliferation, in the particular case of atherosclerosis (see Motterlini and Otterbein, 2010). The first applied patent for the use of CO in medicine was in 2003 (Yale University: WO03094932A1; 2003; for further information on CO-based patents, see Zuckerbraun, 2008; Bannenberg and Vieira, 2009).

Haem oxygenase

HO can be found in two main isoforms: HO-1 or inducible and HO-2 or constitutive. Both respond to stress by increasing their expression or activity respectively (Ryter et al., 2006). A further isoform, HO-3, has been found only in the rat brain (McCoubrey et al., 1997) and not in the human. HO is expressed/activated in response to a wide range of different cellular stress conditions, such as oxidative stress, hyperthermia, hypothermia, ischaemia, hypoxia, hyperoxia, inflammation or exposure to UV light(Gozzelino et al., 2010). HO plays a crucial role in the redox state of the cell and is crucial for cellular maintenance and survival in many organ systems, such as brain (Dore, 2002), heart (Piantadosi et al., 2008), intestine (Nakao et al., 2008), liver (Babu et al., 2007) and lung (Morse et al., 2009). The contribution of HO to the maintenance of both tissue homeostasis and cytoprotection is due to two main actions: (i) the removal of the haem group (originating from dying cells or from haemoglobin following haemorrhage) and (ii) the biological activity of HO products (Grochot-Przeczek *et al.*, 2012). It is worth noting that under stress, haem-containing proteins can release free haem, which becomes a potent oxidising agent through the Fenton reaction. For this reason, the catabolism of free haem by HO is crucial for maintaining tissue homeostasis and cytoprotection (see Gozzelino *et al.*, 2010).

HO and the CNS

In the brain, basal HO-1 expression is low, whereas under stress stimulation, it increases in neuronal, glial and endothelial cells. Likewise, constitutive expression of HO-2 is mainly distributed in mammalian neuraxis, but its expression can also increase following damaging stimuli (Schipper, 2004a,b), such as in hypoxic–ischaemic insult (Sutherland *et al.*, 2009). In several brain pathologies, HO expression and activity can be involved in the modulation of disease development, as well as in the re-establishment of tissue homeostasis.

HO levels as potential biomarkers in the CNS

The use of HO protein as a biomarker for brain damage presents some limitations. Firstly, increased levels of HO-1 in the human serum are not brain-specific and can indicate systemic inflammation and/or tissue damage. Secondly, although it is widely accepted that HO expression is associated with neuroprotection, glial cytoprotection and anti-inflammatory events, HO responds to several different stress stimuli. Likewise, increased expression of HO indicates pathological changes in the tissue. It is, therefore, quite difficult to interpret high-risk changes in the expression of HO and to decide if it is a biomarker of pathological processes or a predictor of a favourable outcome.

For example, the levels of HO-1 in the CSF of infants and children after severe traumatic brain injury are higher (Cousar *et al.*, 2006). Likewise, the level of HO-1 protein is a promising serum biomarker for early detection of Alzheimer's disease (AD), as they seem to increase in patients with AD and



mild cognitive impairment (Mueller et al., 2010). An example showing that HO-1 levels can be used as a diagnostic and prognostic biomarker is during hypothermia treatment following haemorrhagic brain injury in a rat model (Yao et al., 2011). The brain cooling-induced decrease on HO-1 expression was associated with an attenuation of oedema formation and a decrease of haem concentration (Yao et al., 2011). Thus, in these three cases, HO-1 expression is associated with the development of pathology and can be used as a diagnostic biomarker. On the contrary, in experimental models of cardiovascular diseases and cerebral ischaemia, lower levels or deletion of HO-1 and -2 expressions are related to a worse outcome (see the following sections). Therefore, enhanced expression of HO could be associated with a favourable outcome being a prognostic biomarker. In summary, HO-1 and HO-2 levels display a potential and promising diagnostic and prognostic value as biomarkers in humans, although further studies are urgently necessary.

In the following sections, examples relating cerebral pathologies and HO are described in a systematic way.

Cerebrovascular diseases: ischaemia and reperfusion

Cerebral ischaemia is the main cause of brain damage and the third major cause of death in Western societies. In adults, it is mainly due to stroke, whereas in infants, it is caused by perinatal complications, particularly birth asphyxia. Cerebral damage is the result of oxygen and tissue energy depletion, leading to acidosis, exacerbated inflammation, glutamate excitotoxicity, oxidative stress and ultimately neural cell death (Dirnagl *et al.*, 2009).

Increasing data indicate that HO-1 activity is crucial for tissue protection and regeneration following cerebral ischaemia. In humans, there is a long-term increase in the expression of HO-1 following focal cerebral infarctions and traumatic brain injury (Beschorner *et al.*, 2000). On the contrary, in a rat model of transient cerebral ischaemia, reduction of HO-1 expression was associated with more severe neuro-degeneration (Moreira *et al.*, 2007). Protection by ischaemic pre-conditioning (IPC) against permanent ischaemic brain injury is dependent on HO-1 expression, as IPC-promoted neuroprotection was abolished in HO-1 gene-deleted mice (Zeynalov and Dore, 2009; Zeynalov *et al.*, 2009). Likewise, overexpression of HO-1 by adenovirus vector treatment attenuated brain damage after focal cerebral ischaemia in rats (Chao *et al.*, 2013).

Modulation of cerebrovasodilation by HO

In neonates, recurrent seizures may result from meningitis, haemorrhage, asphyxia, and hypoxia or metabolic disorders. Neonatal seizures may promote neuronal damage and susceptibility to epilepsy in survivors. Both HO-1 and HO-2 activities in astrocytes, neurons, endothelial cells and smooth muscle cells (in cerebral vessels) are involved in the modulation of cerebral blood flow and vasodilation during seizures (Parfenova *et al.*, 2003; 2012a; Basuroy *et al.*, 2006; Xi *et al.*, 2011). Moreover, ionotropic glutamate receptors mediate HO activation and endogenous production of CO, which increases cerebral blood flow, essential for maintaining brain homeostasis and neuronal survival during seizures (Parfenova *et al.*, 2012b).

HO and AD

Despite the increasing amount of data demonstrating HO as a widespread cytoprotective enzyme, its homeostatic and neuroprotective role in AD is somewhat controversial. AD is associated with an increased deposition of redox-active iron, chronic oxidative stress and mitochondrial malfunctioning, which are implicated in the development of this pathological disorder. Indeed, in experimental models, glial overexpression of HO-1 promoted mitochondrial oxidative stress (Song *et al.*, 2006) and mediated mitochondrial membrane damage and autophagy in astrocytes (Zukor *et al.*, 2009). Additionally, in mouse brain, long-term overexpression of HO-1 induced toxic tau accumulation (Hui *et al.*, 2011) and increased deposits of glial iron (Song *et al.*, 2012).

On the contrary, HO expression appears to be involved in reduction of brain oxidative stress. In an ageing canine model, which develops cognitive dysfunction and neuropathology similar to those in human AD patients, atorvastatin-induced up-regulation of HO was associated with reduced oxidative stress (Butterfield *et al.*, 2012). In the same canine model, brain oxidative stress biomarkers (protein carbonyl, 3-nitrotyrosine and levels of products of lipid peroxidation) were attenuated following enriched environment-antioxidant-fortified feeding, which was strongly associated with an enhancement of HO-1 protein levels (Opii *et al.*, 2008).

HO suppressor factors, such as α 1-antitrypsin, may also play a role in the development of AD, as Maes *et al.* (2006) have found significantly augmented plasma HO suppressor activity in AD patients, relative to healthy elderly subjects.

As previously mentioned, HO levels were increased in the serum of AD patients, and could be being a potential diagnostic biomarker (Mueller *et al.*, 2010). In addition, HO post-translational modification might also be involved in the development of AD. Barone and colleagues found that HO-1 protein levels were significantly increased in the hippocampus of AD subjects, whereas HO-2 protein levels were significantly decreased in both AD and mild cognitive impairment hippocampi. Serine phosphorylation and increased oxidative, post-translational, modifications of HO-1 were also found in the hippocampus of AD patients (Barone *et al.*, 2012). Controversially, it was also observed that HO-1 protein levels are lower in *post mortem* specimens of CSF (see Schipper 2000).

Thus, HO isoforms and protein post-translational modifications might also play a role in the debate between neuro-protective versus neurotoxic effects of HO activity in AD.

HO and Parkinson's disease (PD)

Oxidative stress, accumulation of Lewy bodies and decrease of mitochondrial complex I activity are common features occurring in nigral dopaminergic neurons (DNs) during pathological development of PD. In *post mortem* human brain specimens collected from PD patients, HO-1 expression was assessed by immunohistochemistry. In the substantia nigra of both PD and control specimens, moderate HO-1 immunore-activity was consistently observed in DNs, while the fraction of GFAP-positive astroglia expressing HO-1 in PD substantia nigra was significantly greater in PD patients (Schipper *et al.*, 1998). Likewise, expression of HO-1 measured by microarray analysis was enhanced following oxidative stress in DNs (Yoo

et al., 2003). Despite the association of HO-1 expression with PD development, HO-1 activity emerges as involved with neuroprotection. For instance, in a rat model of MPP+ (1-methyl-4-phenylpyridinium)-induced PD, local injection of adenovirus containing human HO-1 gene increased the survival rate of DNs and reduced the production of TNF- α (Hung et al., 2008). Using an in vitro model of rat midbrain slice culture, in which DNs were induced to die by IFN-γ/LPS treatment, surviving neurons displayed more robust expression of HO-1, whereas treatment with a HO-1 inhibitor, zinc protoporphyrin IX, increased cell death (Kurauchi et al., 2009). Fibroblast growth factor 9 prevented MPP-induced nigral dopaminergic neuronal death via up-regulation of HO-1 (Huang and Chuang, 2010). In the autosomal recessive form of PD, due to the PINK1 G309D mutation, there is an impairment of HO-1 production in response to oxidative stress (Chien et al., 2011). In addition, HO-1 activity also seems to be associated with modulation of proteasome degradation, whose activity is decreased in patients with PD. Indeed, misfolding proteins promote neuronal toxic stimuli, which induce HO-1 expression, and, in turn, prevent intracellular accumulation of protein aggregates and inclusions in human neuroblastoma M17 cells (Song et al., 2009). Controversially, HO-1 knockout mice treated with MPP i.p. injection for inducing PD presented the same levels of dopaminergic degeneration and severity of gliosis as control animals (Innamorato et al., 2010).

In summary, although HO activity is associated with cytoprotection and neuroprotection, some authors have suggested that it is implicated in neurotoxicity and should be a therapeutic target for chronic neurodegenerative diseases (AD and PD), namely through the prevention of its expression and/or activity for avoiding iron accumulation. Indeed, Schipper et al. (2009) suggested the suppression of glial HO-1 activity as a potential therapeutic strategy for treating AD. Furthermore, the levels of ferritin protein are crucial for maintaining a functional cellular iron storage, whose role must be coupled with HO activity. Ferritin is a very important protein with a dual role of protecting the cell against the oxidative stress caused by free iron, yet allowing access to it. There are two isoforms, L and H, distributed throughout the tissues. L-ferritin has iron nucleation properties and a mutation on this chain leads to iron deposition in cerebellum, basal ganglia and motor cortex, causing an autosomal dominant inherited disorder (neuroferritinopathy) (Lehn et al., 2012). Additionally, H-ferritin mutations lead to a propensity to oxidative stress, notwithstanding normal iron concentration, as the L-ferritin compensates for the loss of H-ferritin. Thus, one can also speculate that, depending upon the ferritin levels and activity, HO could promote cytoprotection or exacerbation of damage. Indeed, Thompson et al. (2003) generated a mouse model for AD and PD, based upon a deficiency on H-ferritin, reinforcing the deleterious role of iron in neurodegenerative diseases. Another important discovery is the existence of mitochondrial ferritin, which is expressed only in the testis and brain (Yang et al., 2013). Despite the lack of data until this date, mitochondrial ferritin is considered to be associated with neuroprotection against neurodegeneration in PD and AD. Thus, the effects of HO on neurodegenerative diseases need to be studied conjointly with ferritin activity.

Neuroinflammation and multiple sclerosis (MS)

During the past decade, several reports have demonstrated that HO activity can also modulate neuroinflammation. HO-1 appears to be involved in the modulation of neuroinflammation because whenever its transcription factor Nrf-2 is knocked out, mice are hypersensitive to LPS-induced neuroinflammation (Innamorato *et al.*, 2008). Still, molecules exerting anti-neuroinflammatory effects, such as dimethyl fumarate (Lin *et al.*, 2011), cyclopentenone prostaglandins (Zhuang *et al.*, 2003b) and 6,4'-dihydroxy-7-methoxyflavanone (Li *et al.*, 2012) act by increasing expression of HO-1.

MS is an autoimmune disease affecting the CNS with inflammatory lesions, demyelination and axonal loss (Fagone et al., 2012). In 2001, the protective and anti-inflammatory role of HO-1 activity in an experimental model of MS, experimental autoimmune encephalomyelitis (EAE) was first shown. Pharmacological induction of HO-1 with haemin effectively inhibited EAE, whereas prevention of HO-1 activity with tin mesoporphyrin exacerbated EAE (Liu et al., 2001). Later, the same effect was demonstrated by genetic inhibition of HO-1, EAE induction in HO-1 knockout mice enhanced CNS demyelination, paralysis and mortality (Chora et al., 2007). Likewise, MS patients present reduced levels of HO-1 expression in peripheral blood mononuclear cells, and during the exacerbation of the disease, there is a significant down regulation of this enzyme (Fagone et al., 2013). In contrast, there is also evidence that overexpression of HO-1 in glial cells was toxic by promoting mitochondrial oxidative stress and damage due to free iron accumulation (Mehindate et al., 2001), and this effect could be reversed by the addition of the iron chelator deferoxamine (Song et al., 2006). Likewise, in astrocytes of spinal cord from MS, patient there were higher levels of HO-1 than in astrocytes from control subjects (Mehindate et al., 2001).

Pain

Carvalho and colleagues proposed the HO-CO-cGMP pathway to be involved in the nociception caused by an acute painful stimulus without inflammation. The administration of pharmacological inhibitor or substrate of HO and soluble guanylyl cyclase (sGC) inhibitor have shown that the antinociceptive action is reduced whenever HO activity is prevented, this effect being dependent upon sGC (Carvalho *et al.*, 2011).

HO in neuroprotection induced by naturally occurring compounds

Epidemiological studies have revealed a reduced incidence of cardiovascular and neurodegeneration risk associated with consumers of specific foods, such berry fruits and red wine. Furthermore, a wide variety of natural compounds extracted from plants or fruits are claimed to promote neuroprotection through modulation of HO-1 expression and/or activity. In 2002, it was first described that in astrocytes, curcumin induces HO-1 expression and activity in a glutathione-independent way (Scapagnini *et al.*, 2002). Since then, several publications have shown, in cultures of neurons and astrocytes, that curcumin protects against inflammation, oxidative



 Table 1

 Natural compounds extracted from plants or fruits that promote neuroprotection through modulation of HO-1 expression and/or activity

Compound	Model	Observations	Reference
Ginkgo biloba	Middle cerebral artery occlusion (transient) and permanent ischaemic stroke	HO-1 KO mice lost beneficial effects	Saleem <i>et al.</i> , 2008; Shah <i>et al.</i> , 2011
Ginkgo biloba	Primary culture of neurons are challenged with oxidative stress and excitotoxicity	HO-1 deleted derived neurons are not protected against cell death	Nada and Shah, 2012
Resveratrol	Ischaemia–reperfusion in rats	Resveratrol attenuated brain tissue damage and increased HO-1 expression	Ren <i>et al.</i> , 2011
Flavanol(-)-epicatechin	Middle cerebral artery occlusion and neuronal culture	Deletion of HO-1 abolished neuroprotective role of this flavanol	Shah et al., 2011
Sevoflurane	Rat model of focal cerebral ischaemia	Induction of HO-1 up-regulation during post-conditioning	Ye <i>et al.</i> , 2012
Triterpenoid	Global ischaemia in rat Focal ischaemia in mice Oxygen-glucose deprivation in neuronal cultures	8 times increase of HO-1 expression in neuronal culture, <i>in vivo</i> enhanced HO-1 expression and reduced neurological dysfunction and infarct volume	Zhang et al., 2012
Octreotide	Middle cerebral artery occlusion	Increased expression of HO-1	Chen et al., 2012

damage and cell death (Table 1). Ginkgo biloba, which is an extract used in traditional Chinese medicine, has been widely described as a neuroprotective substance. In Table 1, there are several examples showing the involvement of HO in Ginkgo biloba-induced neuroprotection using *in vitro* and *in vivo* models. Resveratrol, which is a component of red wine associated with cardioprotection and neuroprotection, was demonstrated to confer its healthy properties by HO-1 activation *in vitro* and *in vivo* (Zhuang *et al.*, 2003a; Sakata *et al.*, 2010), (Ren *et al.*, 2011). Finally, other natural occurring compounds, such as flavanol(–)-epicatechin, sevoflurane, triterpenoid and octreotide, are also implicated in neuroprotection via HO-1 activation (Table 1).

Carbon monoxide and carbon monoxide-releasing molecules (CORMs)

During the past two decades, many biological functions of CO have been described and great efforts are being made to develop its use for human health. The potential clinical application of inhaled carbon monoxide presents several disadvantages: (i) inhaled CO is not tissue specific; (ii) CO gas is, at least partly, delivered in the body through blood plasma flow and carboxyhaemoglobin, leads to partial systemic hypoxia and toxicity; and (iii) the need of hospital facilities with technical devices for CO inhalation and monitoring oxygen blood levels. To overcome these limitations, great efforts have been taken by chemists to create pro-drugs by synthesizing molecules able to deliver CO, which were first called carbon monoxide-releasing molecules - CORMs (Motterlini et al., 2002). Although a large number of CORMs were developed in the last decade, only few of them have shown proven and efficient beneficial biological effects in in vivo and in vitro

systems. Several issues must be overcome in the development of CORMs, namely water insolubility, toxic chemical structures, promotion of high levels of carboxyhaemoglobin and chemical instability (for further review on their development, see Romao et al., 2012). In the particular case of the CNS, the most studied pro-drugs were CORM-A1, CORM-2 and CORM-3. CORM-A1 ([H₃BCO₂]Na₂) is a boranocarbonate molecule (Motterlini et al., 2005), whereas the transition metal-based molecules are CORM-2 [Ru(CO)₃Cl₂]₂, which is a dimer and insoluble in water (Motterlini et al., 2002), and the water-soluble CORM-3 ([Ru(CO)₃Cl(H₂NCH₂CO₂)]) (Clark et al., 2003). Furthermore, in the specific case of experimental cerebral malaria, a new ruthenium-based molecule was tested, ALF 492, presenting CORM-3 structure with methylthiogalactopyranoside ligand (Pena et al., 2012). The molybdenum-based water-soluble molecule ALF 186 was shown to confer neuroprotection (Schallner et al., 2013).

Developing drugs for brain pathologies is highly challenging due to its extreme importance and complexity, as well as due to the presence of blood-brain barrier (BBB), a biological barrier constituted by the endothelial cells of the blood capillaries together with associated astrocytic end-feet processes and perivascular neurons. The BBB serves to isolate the brain and decreases the risk of infection and the entrance of toxins. Although much work has been done on CORMs and the brain, the ability of any CORM to cross the BBB has not been fully clarified, while it is accepted that the released CO gas could cross biological membranes.

CO and the CNS

Exogenous administration of low levels of CO (as CO gas or as CORMs) has been explored as potential therapeutic factor in many different models of brain pathologies (Figure 2), which are described in this section.

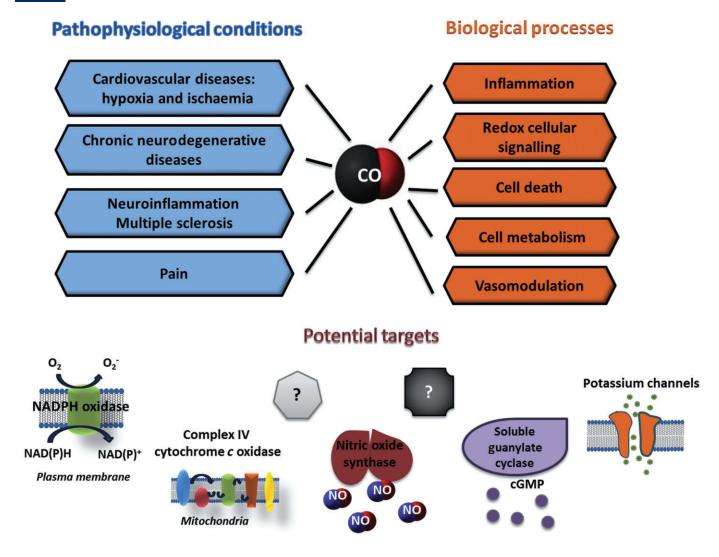


Figure 2Schematic representation of the pathophysiological conditions, biological processes and potential targets for CO.

Cerebrovascular disease

Low levels of inhaled CO were beneficial against cerebral hypoxic and ischaemic insult in experimental rodent models. In mice, CO exposure at 250 ppm for 18 h immediately after permanent middle cerebral artery occlusion (MCAO) decreased the infarct volume by about 30% after 7 days (Wang et al., 2011). Likewise, in a transient MCAO model (90 min of focal ischaemia followed by 48 h of reperfusion), inhalation of 125 ppm of CO immediately at the onset of reperfusion also decreased brain damage by about 30% after 48 h. Inhalation of CO at 250 ppm in the same model and conditions decreased brain damage by around 60%. Interestingly, when the CO inhalation is performed 1 h or 3 h after the reperfusion, there is still reduction in brain damage, by 70 and 30%, respectively (Zeynalov and Dore, 2009). In a rat model of haemorrhagic stroke, when CORM-3 was administered 5 min before or 3 days after, the intracerebral haemorrhage stimulus (injection of collagenase), inflammatory responses were decreased. The opposite effect was achieved when CORM-3 is injected 3 h after the haemorrhagic insult (Yabluchanskiy et al., 2012). Thus, the time window for CO administration is a crucial aspect for its biological functions. In a perinatal rat model of cerebral hypoxia-ischaemia, CO exposure at 250 ppm for 1 h·day⁻¹ on the 3 days prior to the ischaemic insult decreased apoptotic cell death in the hippocampus by 64% (Queiroga et al., 2012b). In another perinatal experimental system, a piglet model of deep hypothermic circulatory arrest, mimicking the open-heart surgery procedures, inhalation of 280 ppm CO for 3 h, 1 day prior to surgery, limited cell death in the neocortex and hippocampus (Mahan et al., 2012). In the in vivo retinal ganglion cell model of ischaemia and reperfusion, CO gas pre-conditioning (Biermann et al., 2010) and postconditioning (Schallner et al., 2012) also promoted neuroprotection. Finally, CORM-A1 (2 mg·kg⁻¹, i.p.) administration 30 min before chemically induced seizures, protected against seizure-induced neonatal vascular injury in newborn piglets (Zimmermann et al., 2007; Parfenova et al., 2012a).



Multiple sclerosis

In the established model of MS, EAE in SJL mice, a prolonged prophylactic treatment with CORM-A1 reduced the incidence of the disease and attenuated the inflammatory infiltrations of the spinal cords (Fagone *et al.*, 2011). Exogenous CO administration (250 ppm) suppressed myelin-reactive immune cell activation within the CNS, contributing to the reduction of autoimmune neuroinflammation impairment (Chora *et al.*, 2007).

Pain

Pain is another aspect where CO has the potential of improving patient quality of life. Hervera and colleagues demonstrated that treating mice with CORM-2 and CORM-3 for 10–20 days following sciatic nerve injury improved the local antinociceptive effects of morphine and significantly reduced the main neuropathic pain symptoms, in a time-dependent manner. Furthermore, this CO effect is due to the reduction of spinal microglial activation and NOS1/NOS2 overexpression (Hervera *et al.*, 2012; 2013a,b).

Brain cells and carbon monoxide: in vitro approaches

Cellular consequences of ischaemia, such as excitotoxicity and oxidative stress, induce cell death and can be mimicked *in vitro*. In primary cultures of cerebellar granular neurons, CO gas limited neuronal cell death via ROS signalling and acting on NOS, sGC and the ATP-dependent mitochondrial K channel (Vieira *et al.*, 2008). Similarly, CO-induced neuroprotection was shown to be dependent upon sGC activity and cGMP production in the SH-SY5Y neuronal cell line and in retinal ganglion cells by using a novel CORM: ALF 186 (Schallner *et al.*, 2013).

Neuroprotection does not concern only neurons. Indeed, one must also target glial cells to achieve complete neuroprotection. The physiological role of astrocyte, microglia, oligodendrocytes and endothelial cells is the maintenance of brain homeostasis, metabolism and neuronal function. Therefore, modulation of glial cell function is crucial for promoting neuroprotection. Likewise, regulation of astrocytic metabolism and prevention of astrocytic apoptosis against oxidative stress is decisive for the maintenance of brain homeostasis. Indeed, CO gas limited astrocytic apoptosis by two distinct ways: (i) direct prevention of mitochondrial membrane permeabilization and the consequent release into the cytosol of pro-apoptotic factors (Queiroga et al., 2010) and (ii) improvement of cellular metabolism and increase of oxidative phosphorylation and mitochondrial population (Almeida et al., 2012).

Furthermore, excessive inflammatory responses can be detrimental and the modulation of inflammation in microglia by CO is very important for the control of neuroinflammation. Many studies of the CO anti-inflammatory effect have been carried out *in vitro* using BV-2 microglial cells. CORM-3 was shown by Bani-Hani and colleagues to decrease NO production and TNF- α release in response to LPS, thrombin and IFN- γ stimuli. inhibition of MAPKs, and PI3K, exacerbated the anti-inflammatory effect of CORM-3. On the opposite, sGC, NOS and HO activity had no influence on the mode of action of CORM-3 (Bani-Hani *et al.*, 2006a,b). Taking all together, the ability of CO to limit inflammatory response

promotes neuronal survival and is important for CO-induced neuroprotection.

Inflammatory brain disease, oxidative stress or excitotoxicity (with excessive glutamate release) might damage cerebral vascular endothelial cells, leading to blood flow dysregulation and permeabilization of the BBB. Parfenova et al demonstrated that cerebral microvascular endothelial cells (CMVECs) contain HO-1 and HO-2 isoforms and their endogenous CO regulates vascular tone in response to glutamate (Parfenova et al., 2001; 2003; Leffler et al., 2011). Likewise, endogenous and exogenous CO prevents endothelial cell death via modulation of Nox4 NADPH activity (Basuroy et al., 2009; 2011) (see the following section). Finally, CORM-A1 prevents BBB dysfunction by limiting glutamate-induced apoptosis and oxidative stress in CMEC (Basuroy et al., 2013). It is possible that the astrocytic end-feet processes and perivascular neurons associated with BBB are the prime targets of CO's effects in the brain.

Pathways involved in CO signalling

Several pathways have been proposed to contribute to the cellular and biochemical mechanisms associated with the biological roles of CO (Figure 2). However, those biochemical pathways and the actual physiological target(s) of CO are still under vigorous discussion (Motterlini and Otterbein, 2010). CO is a rather chemically inert molecule, and in biological systems, it can only bind to transition metals present in several proteins (Boczkowski *et al.*, 2006), thus modulating their activity. In mammals, iron-containing haem-proteins are the most studied and documented targets for CO. Notably, CO can only bind to reduced Fe²⁺, limiting the potential target proteins, in contrast to NO that binds to both Fe²⁺ and Fe³⁺ (Boczkowski *et al.*, 2006).

In the CNS, the pathways and potential targets of CO are still poorly understood with few available data published concerning the mechanisms by which CO confers neuroprotection, anti-neuroinflammation or vasomodulation and this lack is crucial to the development of more scientific research on this subject. This section focuses upon and discusses the existing data about the pathways used by CO in the brain.

sGC and NOS

One of the most studied pathways for CO is the activation of sGC and NOS. Nevertheless, the binding affinity of CO for sGC is still controversial under physiological conditions, as high concentrations of CO are usually required for activating sGC, compared with the much lower levels of NO that are needed for activating sGC. Regarding neuronal cells, activation of sGC and NOS and the respective production of cGMP and NO were shown to be important for CO-induced neuroprotection against excitotoxicity and ischaemic insult (Vieira et al., 2008; Schallner et al., 2013). In a model of permanent ischaemic stroke, the protective role of HO-1 is correlated with higher levels of endothelial NOS expression in the brain (Shah et al., 2011). Likewise, in a neuroinflammatory model, CO regulates inflammation in microgial cells by modulating NO production (Bani-Hani et al., 2006a,b). Still, increased levels of cGMP appeared to be downstream to endogenous

CO production in astrocytes (Imuta *et al.*, 2007), whereas in cerebral microvessels, cGMP signalling appeared to be upstream of CO modulation, because glutamate-induced NOS activation led to CO production via cGMP signalling (Leffler *et al.*, 2005).

Finally, CO appears also to modulate pain through NO signalling. The antinociceptive effects of morphine and agonists of μ -opioid receptors, δ -opioid receptors and cannabinoid CB₂ receptors are enhanced by CO (CORM-2 and CORM-3) in a NO-dependent fashion, during chronic inflammatory and neuropathic pain (Hervera *et al.*, 2013a,b).

Reactive oxygen species (ROS) signalling

It is increasingly accepted in several cell and tissue models that the mediation of CO-induced cytoprotection is via ROS generation and signalling (see Bilban *et al.*, 2008; Queiroga *et al.*, 2012a). At least two cellular proteins are recognized as being directly implicated in cell redox signalling by CO: cytochrome *c* oxidase (mitochondrial respiratory complex IV) and NAD(P)H oxidase (plasmatic membrane).

Cytochrome c oxidase is the main described target for the cytotoxic effects of CO as, by binding to cytochrome c oxidase, CO blocks mitochondrial respiration, promoting cell death (Wainio and Greenless, 1960; Savolainen et al., 1980; Alonso et al., 2003). Furthermore, endogenous CO can also control and inhibit cellular respiration through acting on cytochrome c oxidase (D'Amico et al., 2006). In neural cells, namely astrocytes, low concentrations of CO present a twostep response regarding cytochrome c oxidase activity. During the first minutes following CO treatment, there is a slight decrease in the cytochrome c oxidase activity, while after 30 min (and up to 24 h), specific activity of cytochrome c oxidase increases (Almeida et al., 2012). Thus, these data indicate a direct action of CO on complex IV of mitochondrial respiratory chain and reinforces the hypotheses claiming that ROS production occurs at complex III level, due to electron accumulation whenever complex IV is inhibited. Likewise, in non-synaptic isolated mitochondria from rat brain cortex, CO promoted ROS generation (Queiroga et al., 2010), and the use of β -carotene for limiting ROS levels prevented the anti-apoptotic effect of CO in astrocytes, as well as the CO-induced protection against mitochondrial membrane permeabilization (Queiroga et al., 2010). In primary cultures of cerebellar granular neurons, small amounts of ROS are produced upon CO treatment and, when ROS generation was prevented by butyl-hydroxytoluene, the neuroprotective effect of CO was reversed, indicating the essential role of ROS as signalling factors (Vieira et al., 2008).

In inflammatory brain diseases, NADPH oxidase, particularly its major isoform Nox4, generates ROS, which can initiate both death and survival pathways in TNF- α -challenged CMVECs. Endogenous and exogenous CO limits the production of superoxide anion by Nox4 NADPH, preventing endothelial cell death caused by TNF- α -induced oxidative stress (Basuroy *et al.*, 2009). Nox4 NADPH-derived ROS also initiated a cell survival mechanism, by increasing production of CO by constitutive HO-2 (Basuroy *et al.*, 2011). The ROS-dependent cell survival pathway is mediated by TNF- α , Akt, ERK1/2 and p38 MAPK signalling pathways (Basuroy *et al.*, 2011). Therefore, there might be a feedback control of ROS production regulated by CO, whereby NADPH oxidase pro-

duced ROS that increased CO generation, which, in turn, prevented NADPH oxidase activity, its excessive production of superoxide anion and oxidative stress.

Outside the nervous system, there are other potential pathways for biological CO action, related to ROS signalling and mitochondria. In cardiomyocytes, CO-induced mitochondrial ROS production may control mitochondrial biogenesis, leading to cytoprotection (Suliman et al., 2007a,b). Likewise, in isolated heart mitochondria, CORM-3 limits excessive mitochondrial ROS production and avoids oxidative stress by inducing a mild-uncoupling state, while complex II seems to be the target of CO as inhibition of complex II (malonate addition) reversed the CO-induced augmentation of oxygen consumption and the uncoupling effect (Lo Iacono et al., 2011). In contrast, in a liver system, CO has been described as a cytoprotective molecule by targeting cytochrome P450 and limiting excessive ROS production and oxidative stress-induced cell death. The best-described example is the isoform cytochrome P450 2E1, which is involved in acetaminophen (paracetamol) hepatotoxicity (Gong et al., 2004). Based upon the data derived from other organs and tissues, neuroscientists should explore other potential targets and pathways for the well-accepted beneficial effects of CO in the brain.

CO and potassium channels

In 2003, it was shown by Tang and colleagues that largeconductance calcium-dependent potassium channels (BKca; K_{Ca}1.1; channel nomenclature follows Alexander et al., 2013b) contain a conserved haem-binding sequence motif, which can bind covalently to haem, regulating its channel activity (Tang et al., 2003). One year later, HO-2-derived CO was demonstrated to modulate the BK_{Ca} channels, which are important for sensing oxygen levels (Williams et al., 2004; Jaggar et al., 2005). Furthermore, endogenous CO may modulate cerebral microvasculature by activating these channels (Jaggar et al., 2005). Particularly, astrocytic HO-2-derived CO causes glutamatergic dilation of pial arterioles, by activating smooth muscle cell BK_{Ca} channels (Leffler et al., 2011). Thus, one can speculate that CO binds directly to BK_{Ca} channelbound haem to control dilation and constriction of vasculature (Leffler et al., 2011).

Future challenges for CO administration

The first challenge to the clinical use of CO in cerebral pathologies is the lack of information about its mode of action. Although many different mechanisms for the cellular and biochemical pathways of CO action have been described, the precise underlying signalling mechanisms and the exact molecular target(s) of CO are poorly defined. It is worth noting that elucidating the potential protein targets of CO under physiological conditions is extremely complex as CO might bind to its target on a dynamic and transitory way. Furthermore, CO directly competes with oxygen for binding to proteins; thus, tissue and cellular oxygen levels also influence the system used to study CO targets under physiological conditions. Based on the fact that CO seems to mimic pre-



conditioning, promoting a tissue tolerance state, one might explore the classical activator and transducer factors involved in pre-conditioning and CO. For instance, pre-conditioning stimulus leads to up-regulation of VEGF (Wick *et al.*, 2002; Laudenbach *et al.*, 2007), activation of hypoxia inducible factor (HIF-1) (Ratan *et al.*, 2004; Chu *et al.*, 2010) or expression of erythropoietin (Ruscher *et al.*, 2002), these factors are promising candidates for CO-related pathways. Indeed, in macrophages, CO has been described as stabilizing HIF-1 (Chin *et al.*, 2007).

The second challenge concerning CO administration is achieving the best way to specifically deliver CO to the target tissue, avoiding high concentrations of carboxyhaemoglobin. Many studies have been made to develop CORMs to avoid the systemic toxicity related to carboxyhaemoglobin (see Romao et al., 2012; Zobi, 2013) and there are now CORMs which induce different levels of carboxyhaemoglobin, Whereas exposure to CO gas and CORM-A1 administration induced similar levels of carboxyhaemoglobin (Otterbein et al., 1999; Ryan et al., 2006), very low levels of carboxyhaemoglobin were observed with CORM-3 (Guo et al., 2004). Nevertheless, how and where to give CORMs to deliver CO efficiently is still a matter of intensive research. Likewise, chemical modifications of CORMs are under progress to target these molecules to a specific organ or cell type (Fagone et al., 2012). So far, the best example is ALF 794 which specifically targets the liver against acute injury (Marques et al., 2012). Still, several questions remain unanswered: How is the CORM transported in blood? Does the CORM bind to any protein present in the blood in order to maintain its stability? Does the CORM need to cross the cellular plasma membrane? Is CO actually delivered in the extracellular space, arriving intracellularly by membrane diffusion? It is clear that many further studies are needed before we can know how the existing CORMs act under physiological conditions. Still, the development of new molecules with optimal control of CO delivery (locus and kinetics) is also crucial for the progress of CO as a novel therapeutic agent for medical applications.

Furthermore, in the brain, another vital biological challenge exists in the shape of the BBB. Several brain studies have been performed *in vivo* using CORM-3 and CORM-A1 with promising results. Although it is not precisely confirmed that these CORMs are able to cross the BBB, CO does enter the brain and acts as a cytoprotective molecule (Zimmermann *et al.*, 2007; Parfenova *et al.*, 2012a; Yabluchanskiy *et al.*, 2012).

The time window for CO administration is essential to optimise the outcome and this factor depends upon the pathophysiological situation. Pre-conditioning is one of the processes induced by CO, where CO stimulates endogenous cellular pathways of protection (anti-inflammatory, antiapoptotic, pro-survival, pro-homeostatic etc.) (Bilban et al., 2008; Piantadosi et al., 2008; Queiroga et al., 2012b). In this case, the therapeutic strategy consists of CO administration prior to the injury; for instance, in patients at high risk of developing cerebral ischaemia (before major cardiac surgery, high-risk newborn infants, ageing patients with cardiovascular complications and risk of ischaemic stroke). Moreover, during the development of chronic diseases, such as AD, PD or MS, exogenous CO can be used as a pre-conditioning agent (Fagone et al., 2011). Other evidence shows that CO has beneficial effects in acute injury, suggesting that CO could be

applied after injury, as a post-conditioning strategy, as described in cerebral ischaemia, intracerebral haemorrhage and seizures (Zeynalov and Dore, 2009; Wang *et al.*, 2011; Yabluchanskiy *et al.*, 2012).

Final conclusions

There is good evidence supporting the protective role of CO (and the enzyme catalysing its biosynthesis, HO) in the CNS in the context of several pathologies, including cerebrovascular diseases, neuroinflammation, MS, pain, AD and PD.

Essential to the therapeutic use of CO is further development of sources of CO, other than CO gas, to overcome the problem of carboxyhaemoglobin toxicity. CORMs have been increasingly used with successful and interesting results. Nevertheless, it was inhaled CO that was first proved to be safe and tolerable in humans.

Independent of the route of administration and regardless of the cell type, CO appears to modulate several important cellular enzyme systems, including cytochrome c oxidase, NOS, sGC and NADPH oxidase. Other targets, such as ROS signalling and mitochondria, are also significant components of the actions of CO. Nevertheless, further research is urgently needed to define more precisely the biological target(s) and pathways of this gasotransmitter. In conclusion, CO has travelled far, from being an invisible enemy to becoming a possible therapeutic solution.

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Conflict of interest

There is no conflict of interest.

References

Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Spedding M *et al.* (2013a). The Concise Guide to PHARMACOLOGY 2013/14: Enzymes. Br J Pharmacol 170: 1797–1867.

Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Catterall WA *et al.* (2013b). The Concise Guide to PHARMACOLOGY 2013/14: Ion Channels. Br J Pharmacol 170: 1607–1651.

Almeida SS, Queiroga CS, Sousa MF, Alves PM, Vieira HL (2012). Carbon monoxide modulates apoptosis by reinforcing oxidative metabolism in astrocytes: role of BCL-2. J Biol Chem 287: 10761–10770.

Alonso JR, Cardellach F, Lopez S, Casademont J, Miro O (2003). Carbon monoxide specifically inhibits cytochrome c oxidase of human mitochondrial respiratory chain. Pharmacol Toxicol 93: 142–146.

BJP C S F Queiroga et al.

Babu AN, Damle SS, Moore EE, Ao L, Song Y, Johnson JL *et al*. (2007). Hemoglobin-based oxygen carrier induces hepatic heme oxygenase 1 expression in Kupffer cells. Surgery 142: 289–294.

Bani-Hani MG, Greenstein D, Mann BE, Green CJ, Motterlini R (2006a). A carbon monoxide-releasing molecule (CORM-3) attenuates lipopolysaccharide- and interferon-gamma-induced inflammation in microglia. Pharmacol Rep 58: 132–144.

Bani-Hani MG, Greenstein D, Mann BE, Green CJ, Motterlini R (2006b). Modulation of thrombin-induced neuroinflammation in BV-2 microglia by carbon monoxide-releasing molecule 3. J Pharmacol Exp Ther 318: 1315–1322.

Bannenberg GL, Vieira HL (2009). Therapeutic applications of the gaseous mediators carbon monoxide and hydrogen sulfide. Expert Opin Ther Pat 19: 663–682.

Barone E, Di Domenico F, Sultana R, Coccia R, Mancuso C, Perluigi M *et al.* (2012). Heme oxygenase-1 posttranslational modifications in the brain of subjects with Alzheimer disease and mild cognitive impairment. Free Radic Biol Med 52: 2292–2301.

Basuroy S, Bhattacharya S, Tcheranova D, Qu Y, Regan RF, Leffler CW *et al.* (2006). HO-2 provides endogenous protection against oxidative stress and apoptosis caused by TNF-alpha in cerebral vascular endothelial cells. Am J Physiol Cell Physiol 291: C897–C908.

Basuroy S, Bhattacharya S, Leffler CW, Parfenova H (2009). Nox4 NADPH oxidase mediates oxidative stress and apoptosis caused by TNF-alpha in cerebral vascular endothelial cells. Am J Physiol Cell Physiol 296: C422–C432.

Basuroy S, Tcheranova D, Bhattacharya S, Leffler CW, Parfenova H (2011). Nox4 NADPH oxidase-derived reactive oxygen species, via endogenous carbon monoxide, promote survival of brain endothelial cells during TNF-alpha-induced apoptosis. Am J Physiol Cell Physiol 300: C256–C265.

Basuroy S, Leffler CW, Parfenova H (2013). CORM-A1 prevents blood-brain barrier dysfunction caused by ionotropic glutamate receptor-mediated endothelial oxidative stress and apoptosis. Am J Physiol Cell Physiol 304: C1105–C1115.

Bernard C (1857) Lecons sur les Effets des Substaces Toxiques et Medicamenteuses. J-B Bailliere et Fils: Paris.

Beschorner R, Adjodah D, Schwab JM, Mittelbronn M, Pedal I, Mattern R *et al.* (2000). Long-term expression of heme oxygenase-1 (HO-1, HSP-32) following focal cerebral infarctions and traumatic brain injury in humans. Acta Neuropathol 100: 377–384.

Biermann J, Lagreze WA, Dimitriu C, Stoykow C, Goebel U (2010). Preconditioning with inhalative carbon monoxide protects rat retinal ganglion cells from ischemia/reperfusion injury. Invest Ophthalmol Vis Sci 51: 3784–3791.

Bilban M, Haschemi A, Wegiel B, Chin BY, Wagner O, Otterbein LE (2008). Heme oxygenase and carbon monoxide initiate homeostatic signaling. J Mol Med 86: 267–279.

Boczkowski J, Poderoso JJ, Motterlini R (2006). CO-metal interaction: vital signaling from a lethal gas. Trends Biochem Sci 31: 614–621.

Brouard S, Otterbein LE, Anrather J, Tobiasch E, Bach FH, Choi AM *et al.* (2000). Carbon monoxide generated by heme oxygenase 1 suppresses endothelial cell apoptosis. J Exp Med 192: 1015–1026.

Butterfield DA, Barone E, Di Domenico F, Cenini G, Sultana R, Murphy MP *et al.* (2012). Atorvastatin treatment in a dog preclinical model of Alzheimer's disease leads to up-regulation of haem oxygenase-1 and is associated with reduced oxidative stress in brain. Int J Neuropsychopharmacol 15: 981–987.

Carvalho PG, Branco LG, Panissi CR (2011). Involvement of the heme oxygenase-carbon monoxide-cGMP pathway in the nociception induced by acute painful stimulus in rats. Brain Res 1385: 107–113.

Chao XD, Ma YH, Luo P, Cao L, Lau WB, Zhao BC *et al.* (2013). Up-regulation of heme oxygenase-1 attenuates brain damage after cerebral ischemia via simultaneous inhibition of superoxide production and preservation of NO bioavailability. Exp Neurol 239: 163–169.

Chen L, Wang L, Zhang X, Cui L, Xing Y, Dong L *et al.* (2012). The protection by octreotide against experimental ischemic stroke: up-regulated transcription factor Nrf2, HO-1 and down-regulated NF-kappaB expression. Brain Res 1475: 80–87.

Chien WL, Lee TR, Hung SY, Kang KH, Lee MJ, Fu WM (2011). Impairment of oxidative stress-induced heme oxygenase-1 expression by the defect of Parkinson-related gene of PINK1. J Neurochem 117: 643–653.

Chin BY, Jiang G, Wegiel B, Wang HJ, Macdonald T, Zhang XC *et al.* (2007). Hypoxia-inducible factor 1alpha stabilization by carbon monoxide results in cytoprotective preconditioning. Proc Natl Acad Sci U S A 104: 5109–5114.

Chora AA, Fontoura P, Cunha A, Pais TF, Cardoso S, Ho PP *et al.* (2007). Heme oxygenase-1 and carbon monoxide suppress autoimmune neuroinflammation. J Clin Invest 117: 438–447.

Chu PW, Beart PM, Jones NM (2010). Preconditioning protects against oxidative injury involving hypoxia-inducible factor-1 and vascular endothelial growth factor in cultured astrocytes. Eur J Pharmacol 633: 24–32.

Clark JE, Naughton P, Shurey S, Green CJ, Johnson TR, Mann BE *et al.* (2003). Cardioprotective actions by a water-soluble carbon monoxide-releasing molecule. Circ Res 93: e2–e8.

Cousar JL, Lai Y, Marco CD, Bayir H, Adelson PD, Janesko-Feldman KL *et al.* (2006). Heme oxygenase 1 in cerebrospinal fluid from infants and children after severe traumatic brain injury. Dev Neurosci 28: 342–347.

D'Amico G, Lam F, Hagen T, Moncada S (2006). Inhibition of cellular respiration by endogenously produced carbon monoxide. J Cell Sci 119: 2291–2298.

Dirnagl U, Becker K, Meisel A (2009). Preconditioning and tolerance against cerebral ischaemia: from experimental strategies to clinical use. Lancet Neurol 8: 398–412.

Dore S (2002). Decreased activity of the antioxidant heme oxygenase enzyme: implications in ischemia and in Alzheimer's disease. Free Radic Biol Med 32: 1276–1282.

Fagone P, Mangano K, Quattrocchi C, Motterlini R, Di Marco R, Magro G *et al.* (2011). Prevention of clinical and histological signs of proteolipid protein (PLP)-induced experimental allergic encephalomyelitis (EAE) in mice by the water-soluble carbon monoxide-releasing molecule (CORM)-A1. Clin Exp Immunol 163: 368–374.

Fagone P, Mangano K, Coco M, Perciavalle V, Garotta G, Romao CC *et al.* (2012). Therapeutic potential of carbon monoxide in multiple sclerosis. Clin Exp Immunol 167: 179–187.

Fagone P, Patti F, Mangano K, Mammana S, Coco M, Touil-Boukoffa C *et al.* (2013). Heme oxygenase-1 expression in peripheral blood mononuclear cells correlates with disease activity in multiple sclerosis. J Neuroimmunol 261: 82–86.

Gong P, Cederbaum AI, Nieto N (2004). Heme oxygenase-1 protects HepG2 cells against cytochrome P450 2E1-dependent toxicity. Free Radic Biol Med 36: 307–318.



Gozzelino R, Jeney V, Soares MP (2010). Mechanisms of cell protection by heme oxygenase-1. Annu Rev Pharmacol Toxicol 50: 323–354.

Grochot-Przeczek A, Dulak J, Jozkowicz A (2012). Haem oxygenase-1: non-canonical roles in physiology and pathology. Clin Sci (Lond) 122: 93–103.

Guo Y, Stein AB, Wu WJ, Tan W, Zhu X, Li QH *et al.* (2004). Administration of a CO-releasing molecule at the time of reperfusion reduces infarct size *in vivo*. Am J Physiol Heart Circ Physiol 286: H1649–H1653.

Haldane J (1895). The action of carbonic oxide on man. J Physiol 18: 430-462.

Harbin TJ, Benignus VA, Muller KE, Barton CN (1988). The effects of low-level carbon monoxide exposure upon evoked cortical potentials in young and elderly men. Neurotoxicol Teratol 10: 93–100.

Hervera A, Leanez S, Negrete R, Motterlini R, Pol O (2012). Carbon monoxide reduces neuropathic pain and spinal microglial activation by inhibiting nitric oxide synthesis in mice. PLoS ONE 7: e43693.

Hervera A, Gou G, Leanez S, Pol O (2013a). Effects of treatment with a carbon monoxide-releasing molecule and a heme oxygenase 1 inducer in the antinociceptive effects of morphine in different models of acute and chronic pain in mice. Psychopharmacology (Berl) 228: 463–477.

Hervera A, Leanez S, Motterlini R, Pol O (2013b). Treatment with carbon monoxide-releasing molecules and an HO-1 inducer enhances the effects and expression of micro-opioid receptors during neuropathic pain. Anesthesiology 118: 1180–1197.

Huang JY, Chuang JI (2010). Fibroblast growth factor 9 upregulates heme oxygenase-1 and gamma-glutamylcysteine synthetase expression to protect neurons from 1-methyl-4-phenylpyridinium toxicity. Free Radic Biol Med 49: 1099–1108.

Hui Y, Wang D, Li W, Zhang L, Jin J, Ma N *et al.* (2011). Long-term overexpression of heme oxygenase 1 promotes tau aggregation in mouse brain by inducing tau phosphorylation. J Alzheimers Dis 26: 299–313

Hung SY, Liou HC, Kang KH, Wu RM, Wen CC, Fu WM (2008). Overexpression of heme oxygenase-1 protects dopaminergic neurons against 1-methyl-4-phenylpyridinium-induced neurotoxicity. Mol Pharmacol 74: 1564–1575.

Imuta N, Hori O, Kitao Y, Tabata Y, Yoshimoto T, Matsuyama T *et al.* (2007). Hypoxia-mediated induction of heme oxygenase type I and carbon monoxide release from astrocytes protects nearby cerebral neurons from hypoxia-mediated apoptosis. Antioxid Redox Signal 9: 543–552.

Innamorato NG, Rojo AI, Garcia-Yague AJ, Yamamoto M, De Ceballos ML, Cuadrado A (2008). The transcription factor Nrf2 is a therapeutic target against brain inflammation. J Immunol 181: 680–689.

Innamorato NG, Jazwa A, Rojo AI, Garcia C, Fernandez-Ruiz J, Grochot-Przeczek A *et al.* (2010). Different susceptibility to the Parkinson's toxin MPTP in mice lacking the redox master regulator Nrf2 or its target gene heme oxygenase-1. PLoS ONE 5: e11838.

Jaggar JH, Li A, Parfenova H, Liu J, Umstot ES, Dopico AM *et al.* (2005). Heme is a carbon monoxide receptor for large-conductance Ca²⁺-activated K+ channels. Circ Res 97: 805–812.

Kurauchi Y, Hisatsune A, Isohama Y, Katsuki H (2009). Nitric oxide-cyclic GMP signaling pathway limits inflammatory

degeneration of midbrain dopaminergic neurons: cell type-specific regulation of heme oxygenase-1 expression. Neuroscience 158: 856–866.

Laudenbach V, Fontaine RH, Medja F, Carmeliet P, Hicklin DJ, Gallego J *et al.* (2007). Neonatal hypoxic preconditioning involves vascular endothelial growth factor. Neurobiol Dis 26: 243–252.

Leffler CW, Balabanova L, Fedinec AL, Parfenova H (2005). Nitric oxide increases carbon monoxide production by piglet cerebral microvessels. Am J Physiol Heart Circ Physiol 289: H1442–H1447.

Leffler CW, Parfenova H, Jaggar JH (2011). Carbon monoxide as an endogenous vascular modulator. Am J Physiol Heart Circ Physiol 301: H1–H11.

Lehn A, Boyle R, Brown H, Airey C, Mellick G (2012). Neuroferritinopathy. Parkinsonism Relat Disord 18: 909–915.

Li B, Lee DS, Jeong GS, Kim YC (2012). Involvement of heme oxygenase-1 induction in the cytoprotective and immunomodulatory activities of

6,4'-dihydroxy-7-methoxyflavanone in murine hippocampal and microglia cells. Eur J Pharmacol 674: 153–162.

Lin SX, Lisi L, Dello Russo C, Polak PE, Sharp A, Weinberg G *et al.* (2011). The anti-inflammatory effects of dimethyl fumarate in astrocytes involve glutathione and haem oxygenase-1. ASN Neuro 3: 75–84.

Liu Y, Zhu B, Luo L, Li P, Paty DW, Cynader MS (2001). Heme oxygenase-1 plays an important protective role in experimental autoimmune encephalomyelitis. Neuroreport 12: 1841–1845.

Lo Iacono L, Boczkowski J, Zini R, Salouage I, Berdeaux A, Motterlini R *et al.* (2011). A carbon monoxide-releasing molecule (CORM-3) uncouples mitochondrial respiration and modulates the production of reactive oxygen species. Free Radic Biol Med 50: 1556–1564.

Maes OC, Kravitz S, Mawal Y, Su H, Liberman A, Mehindate K *et al.* (2006). Characterization of alpha1-antitrypsin as a heme oxygenase-1 suppressor in Alzheimer plasma. Neurobiol Dis 24: 89–100.

Mahan VL, Zurakowski D, Otterbein LE, Pigula FA (2012). Inhaled carbon monoxide provides cerebral cytoprotection in pigs. PLoS ONE 7: e41982.

Marques AR, Kromer L, Gallo DJ, Penacho N, Rodrigues SS, Seixas JD *et al.* (2012). Generation of carbon monoxide releasing molecules (CO-RMs) as drug candidates for the treatment of acute liver injury: targeting of CO-RMs to the liver. Organometallics 31: 5810–5822.

McCoubrey WK Jr, Huang TJ, Maines MD (1997). Isolation and characterization of a cDNA from the rat brain that encodes hemoprotein heme oxygenase-3. Eur J Biochem 247: 725–732.

Mehindate K, Sahlas DJ, Frankel D, Mawal Y, Liberman A, Corcos J *et al.* (2001). Proinflammatory cytokines promote glial heme oxygenase-1 expression and mitochondrial iron deposition: implications for multiple sclerosis. J Neurochem 77: 1386–1395.

Moreira TJ, Cebere A, Cebers G, Ostenson CG, Efendic S, Liljequist S (2007). Reduced HO-1 protein expression is associated with more severe neurodegeneration after transient ischemia induced by cortical compression in diabetic Goto-Kakizaki rats. J Cereb Blood Flow Metab 27: 1710–1723.

Morse D, Lin L, Choi AM, Ryter SW (2009). Heme oxygenase-1, a critical arbitrator of cell death pathways in lung injury and disease. Free Radic Biol Med 47: 1–12.

Motterlini R, Otterbein LE (2010). The therapeutic potential of carbon monoxide. Nat Rev Drug Discov 9: 728–743.

Motterlini R, Clark JE, Foresti R, Sarathchandra P, Mann BE, Green CJ (2002). Carbon monoxide-releasing molecules: characterization of biochemical and vascular activities. Circ Res 90: E17–E24.

Motterlini R, Mann BE, Foresti R (2005). Therapeutic applications of carbon monoxide-releasing molecules. Expert Opin Investig Drugs 14: 1305–1318.

Mueller C, Zhou W, Vanmeter A, Heiby M, Magaki S, Ross MM *et al.* (2010). The heme degradation pathway is a promising serum biomarker source for the early detection of Alzheimer's disease. J Alzheimers Dis 19: 1081–1091.

Nada SE, Shah ZA (2012). Preconditioning with Ginkgo biloba (EGb 761(R)) provides neuroprotection through HO1 and CRMP2. Neurobiol Dis 46: 180–189.

Nakao A, Kaczorowski DJ, Sugimoto R, Billiar TR, Mccurry KR (2008). Application of heme oxygenase-1, carbon monoxide and biliverdin for the prevention of intestinal ischemia/reperfusion injury. J Clin Biochem Nutr 42: 78–88.

Opii WO, Joshi G, Head E, Milgram NW, Muggenburg BA, Klein JB *et al.* (2008). Proteomic identification of brain proteins in the canine model of human aging following a long-term treatment with antioxidants and a program of behavioral enrichment: relevance to Alzheimer's disease. Neurobiol Aging 29: 51–70.

Otterbein LE, Mantell LL, Choi AM (1999). Carbon monoxide provides protection against hyperoxic lung injury. Am J Physiol 276: L688–L694.

Otterbein LE, Bach FH, Alam J, Soares M, Tao Lu H, Wysk M *et al.* (2000). Carbon monoxide has anti-inflammatory effects involving the mitogen-activated protein kinase pathway. Nat Med 6: 422–428.

Parfenova H, NEff RA 3rd, Alonso JS, Shlopov BV, Jamal CN, Sarkisova SA *et al.* (2001). Cerebral vascular endothelial heme oxygenase: expression, localization, and activation by glutamate. Am J Physiol Cell Physiol 281: C1954–C1963.

Parfenova H, Fedinec A, Leffler CW (2003). Ionotropic glutamate receptors in cerebral microvascular endothelium are functionally linked to heme oxygenase. J Cereb Blood Flow Metab 23: 190–197.

Parfenova H, Leffler CW, Basuroy S, Liu J, Fedinec AL (2012a). Antioxidant roles of heme oxygenase, carbon monoxide, and bilirubin in cerebral circulation during seizures. J Cereb Blood Flow Metab 32: 1024–1034.

Parfenova H, Tcheranova D, Basuroy S, FedineC AL, Liu J, Leffler CW (2012b). Functional role of astrocyte glutamate receptors and carbon monoxide in cerebral vasodilation response to glutamate. Am J Physiol Heart Circ Physiol 302: H2257–H2266.

Pena AC, Penacho N, Mancio-Silva L, Neres R, Seixas JD, Fernandes AC *et al.* (2012). A novel carbon monoxide-releasing molecule fully protects mice from severe malaria. Antimicrob Agents Chemother 56: 1281–1290.

Piantadosi CA (2002). Biological chemistry of carbon monoxide. Antioxid Redox Signal 4: 259–270.

Piantadosi CA, Carraway MS, Babiker A, Suliman HB (2008). Heme oxygenase-1 regulates cardiac mitochondrial biogenesis via Nrf2-mediated transcriptional control of nuclear respiratory factor-1. Circ Res 103: 1232–1240.

Queiroga CS, Almeida AS, Martel C, Brenner C, Alves PM, Vieira HL (2010). Glutathionylation of adenine nucleotide translocase induced by carbon monoxide prevents mitochondrial membrane permeabilization and apoptosis. J Biol Chem 285: 17077–17088.

Queiroga CS, Almeida AS, Vieira HL (2012a). Carbon monoxide targeting mitochondria. Biochem Res Int 2012: 749845.

Queiroga CSF, Tomasi S, Widerøe M, Alves PM, Vercelli A, Veira HLA (2012b). Preconditioning triggered by carbon monoxide (CO) provides neuronal protection following perinatal hypoxia-ischemia. PLoS ONE 7: e42632.

Ratan RR, Siddiq A, Aminova L, Lange PS, Langley B, Ayoub I *et al.* (2004). Translation of ischemic preconditioning to the patient: prolyl hydroxylase inhibition and hypoxia inducible factor-1 as novel targets for stroke therapy. Stroke 35: 2687–2689.

Ren J, Fan C, Chen N, Huang J, Yang Q (2011). Resveratrol pretreatment attenuates cerebral ischemic injury by upregulating expression of transcription factor Nrf2 and HO-1 in rats. Neurochem Res 36: 2352–2362.

Romao CC, Blattler WA, Seixas JD, Bernardes GJ (2012). Developing drug molecules for therapy with carbon monoxide. Chem Soc Rev 41: 3571–3583.

Ruscher K, Freyer D, Karsch M, Isaev N, Megow D, Sawitzki B *et al.* (2002). Erythropoietin is a paracrine mediator of ischemic tolerance in the brain: evidence from an *in vitro* model. J Neurosci 22: 10291–10301.

Ryan MJ, Jernigan NL, Drummond HA, Mclemore GR Jr, Rimoldi JM, Poreddy SR *et al.* (2006). Renal vascular responses to CORM-A1 in the mouse. Pharmacol Res 54: 24–29.

Ryter SW, Alam J, Choi AM (2006). Heme oxygenase-1/carbon monoxide: from basic science to therapeutic applications. Physiol Rev 86: 583–650.

Sakata Y, Zhuang H, Kwansa H, Koehler RC, Dore S (2010). Resveratrol protects against experimental stroke: putative neuroprotective role of heme oxygenase 1. Exp Neurol 224: 325–329.

Saleem S, Zhuang H, Biswal S, Christen Y, Dore S (2008). Ginkgo biloba extract neuroprotective action is dependent on heme oxygenase 1 in ischemic reperfusion brain injury. Stroke 39: 3389–3396.

Sammut IA, Foresti R, Clark JE, Exon DJ, Vesely MJ, Sarathchandra P *et al.* (1998). Carbon monoxide is a major contributor to the regulation of vascular tone in aortas expressing high levels of haeme oxygenase-1. Br J Pharmacol 125: 1437–1444.

Savolainen H, Kurppa K, Tenhunen R, Kivisto H (1980). Biochemical effects of carbon monoxide poisoning in rat brain with special reference to blood carboxyhemoglobin and cerebral cytochrome oxidase activity. Neurosci Lett 19: 319–323.

Scapagnini G, Foresti R, Calabrese V, Giuffrida Stella AM, Green CJ, Motterlini R (2002). Caffeic acid phenethyl ester and curcumin: a novel class of heme oxygenase-1 inducers. Mol Pharmacol 61: 554–561.

Schallner N, Fuchs M, Schwer CI, Loop T, Buerkle H, Lagreze WA *et al.* (2012). Postconditioning with inhaled carbon monoxide counteracts apoptosis and neuroinflammation in the ischemic rat retina. PLoS ONE 7: e46479.

Schallner N, Romao CC, Biermann J, Lagreze WA, Otterbein LE, Buerkle H *et al.* (2013). Carbon monoxide abrogates ischemic insult to neuronal cells via the soluble guanylate cyclase-cGMP pathway. PLoS ONE 8: e60672.

Schipper HM (2000). Heme oxygenase-1: role in brain aging and neurodegeneration. Exp Gerontol 35: 821–830.

Schipper HM (2004a). Heme oxygenase-1: transducer of pathological brain iron sequestration under oxidative stress. Ann N Y Acad Sci 1012: 84–93.

Schipper HM (2004b). Heme oxygenase expression in human central nervous system disorders. Free Radic Biol Med 37: 1995–2011



Schipper HM, Liberman A, Stopa EG (1998). Neural heme oxygenase-1 expression in idiopathic Parkinson's disease. Exp Neurol 150: 60–68.

Schipper HM, Gupta A, Szarek WA (2009). Suppression of glial HO-1 activity as a potential neurotherapeutic intervention in AD. Curr Alzheimer Res 6: 424–430.

Shah ZA, Nada SE, Dore S (2011). Heme oxygenase 1, beneficial role in permanent ischemic stroke and in Gingko biloba (EGb 761) neuroprotection. Neuroscience 180: 248–255.

Sjostrand T (1949). Endogenous formation of carbon monoxide in man. Nature 164: 580.

Song W, Su H, Song S, Paudel HK, Schipper HM (2006). Over-expression of heme oxygenase-1 promotes oxidative mitochondrial damage in rat astroglia. J Cell Physiol 206: 655–663.

Song W, Patel A, Qureshi HY, Han D, Schipper HM, Paudel HK (2009). The Parkinson disease-associated A30P mutation stabilizes alpha-synuclein against proteasomal degradation triggered by heme oxygenase-1 over-expression in human neuroblastoma cells. J Neurochem 110: 719–733.

Song W, Zukor H, Lin SH, Liberman A, Tavitian A, Mui J *et al.* (2012). Unregulated brain iron deposition in transgenic mice over-expressing HMOX1 in the astrocytic compartment. J Neurochem 123: 325–336.

Suliman HB, Carraway MS, Ali AS, Reynolds CM, Welty-Wolf KE, Piantadosi CA (2007a). The CO/HO system reverses inhibition of mitochondrial biogenesis and prevents murine doxorubicin cardiomyopathy. J Clin Invest 117: 3730–3741.

Suliman HB, Carraway MS, Tatro LG, Piantadosi CA (2007b). A new activating role for CO in cardiac mitochondrial biogenesis. J Cell Sci 120: 299–308.

Sutherland BA, Rahman RM, Clarkson AN, Shaw OM, Nair SM, Appleton I (2009). Cerebral heme oxygenase 1 and 2 spatial distribution is modulated following injury from hypoxia-ischemia and middle cerebral artery occlusion in rats. Neurosci Res 65: 326–334.

Tang XD, Xu R, Reynolds MF, Garcia ML, Heinemann SH, Hoshi T (2003). Haem can bind to and inhibit mammalian calcium-dependent Slo1 BK channels. Nature 425: 531–535.

Tenhunen R, Marver HS, Schmid R (1968). The enzymatic conversion of heme to bilirubin by microsomal heme oxygenase. Proc Natl Acad Sci U S A 61:748-755.

Thompson K, Menzies S, Muckenthaler M, Torti FM, Wood T, Torti SV *et al.* (2003). Mouse brains deficient in H-ferritin have normal iron concentration but a protein profile of iron deficiency and increased evidence of oxidative stress. J Neurosci Res 71: 46–63.

Verma A, Hirsch DJ, Glatt CE, Ronnett GV, Snyder SH (1993). Carbon monoxide: a putative neural messenger. Science 259: 381–384.

Vieira HL, Queiroga CS, Alves PM (2008). Preconditioning induced by carbon monoxide provides neuronal protection against apoptosis. J Neurochem 107: 375–384.

Wainio WW, Greenless J (1960). Complexes of cytochrome c oxidase with cyanide and carbon monoxide. Arch Biochem Biophys 90: 18–21.

Wang B, Cao W, Biswal S, Dore S (2011). Carbon monoxide-activated Nrf2 pathway leads to protection against permanent focal cerebral ischemia. Stroke 42: 2605–2610.

Wick A, Wick W, Waltenberger J, Weller M, Dichgans J, Schulz JB (2002). Neuroprotection by hypoxic preconditioning requires

sequential activation of vascular endothelial growth factor receptor and Akt. J Neurosci 22: 6401–6407.

Williams SE, Wootton P, Mason HS, Bould J, Iles DE, Riccardi D *et al.* (2004). Hemoxygenase-2 is an oxygen sensor for a calcium-sensitive potassium channel. Science 306: 2093–2097.

Xi Q, Tcheranova D, Basuroy S, Parfenova H, Jaggar JH, Leffler CW (2011). Glutamate-induced calcium signals stimulate CO production in piglet astrocytes. Am J Physiol Heart Circ Physiol 301: H428–H433.

Yabluchanskiy A, Sawle P, Homer-Vanniasinkam S, Green CJ, Foresti R, Motterlini R (2012). CORM-3, a carbon monoxide-releasing molecule, alters the inflammatory response and reduces brain damage in a rat model of hemorrhagic stroke. Crit Care Med 40: 544–552.

Yang H, Yang M, Guan H, Liu Z, Zhao S, Takeuchi S *et al.* (2013). Mitochondrial ferritin in neurodegenerative diseases. Neurosci Res 77: 1–7.

Yao C, Wei G, Lu XC, Yang W, Tortella FC, Dave JR (2011). Selective brain cooling in rats ameliorates intracerebral hemorrhage and edema caused by penetrating brain injury: possible involvement of heme oxygenase-1 expression. J Neurotrauma 28: 1237–1245.

Ye Z, Guo Q, Xia P, Wang N, Wang E, Yuan Y (2012). Sevoflurane postconditioning involves an up-regulation of HIF-1alpha and HO-1 expression via PI3K/Akt pathway in a rat model of focal cerebral ischemia. Brain Res 1463: 63–74.

Yoo MS, Chun HS, Son JJ, Degiorgio LA, Kim DJ, Peng C *et al.* (2003). Oxidative stress regulated genes in nigral dopaminergic neuronal cells: correlation with the known pathology in Parkinson's disease. Brain Res Mol Brain Res 110: 76–84.

Zeynalov E, Dore S (2009). Low doses of carbon monoxide protect against experimental focal brain ischemia. Neurotox Res 15: 133–137.

Zeynalov E, Shah ZA, Li RC, Dore S (2009). Heme oxygenase 1 is associated with ischemic preconditioning-induced protection against brain ischemia. Neurobiol Dis 35: 264–269.

Zhang F, Wang S, Zhang M, Weng Z, Li P, Gan Y *et al.* (2012). Pharmacological induction of heme oxygenase-1 by a triterpenoid protects neurons against ischemic injury. Stroke 43: 1390–1397.

Zhuang H, Kim YS, Koehler RC, Dore S (2003a). Potential mechanism by which resveratrol, a red wine constituent, protects neurons. Ann N Y Acad Sci 993: 276–286, discussion 287–8.

Zhuang H, Kim YS, Namiranian K, Dore S (2003b). Prostaglandins of J series control heme oxygenase expression: potential significance in modulating neuroinflammation. Ann N Y Acad Sci 993: 208–216, discussion 287–8.

Zimmermann A, Leffler CW, Tcheranova D, Fedinec AL, Parfenova H (2007). Cerebroprotective effects of the CO-releasing molecule CORM-A1 against seizure-induced neonatal vascular injury. Am J Physiol Heart Circ Physiol 293: H2501–H2507.

Zobi F (2013). CO and CO-releasing molecules in medicinal chemistry. Future Med Chem 5: 175–188.

Zuckerbraun BS (2008). Therapeutic delivery of carbon monoxide: WO2008/003953. Expert Opin Ther Pat 18: 1321–1325.

Zukor H, Song W, Liberman A, Mui J, Vali H, Fillebeen C *et al.* (2009). HO-1-mediated macroautophagy: a mechanism for unregulated iron deposition in aging and degenerating neural tissues. J Neurochem 109: 776–791.